Efficacy of Crude Leaf Extracts of *Aloe secundiflora* on Selected Enteric Bacterial Pathogens and *Candida albicans*

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Introduction

Medicinal plants are used by almost 80% of the world's population for their basic health care because of their low cost and ease in availability [1]. From the dawn of civilization, people have developed great interest in plant based drugs and pharmaceutical products [1]. In the last few decades many bacterial organisms have continued to show increasing resistance against current antimicrobial agents [2]. Herbal drugs made from medicinal plants have been used from ancient times to treat various diseases and their antimicrobial properties make them a rich source of many potent drugs [3]. The use of herbal medicinal plants has always played a positive role in the control or prevention of diseases such as diabetes, heart disorders and various cancers [4]. Some medicinal plants have been used in production of various drugs singly or in combination and even as principal raw material for the production of other conventional medicines [2]. Aloe extracts have been used for many centuries for their curative and therapeutic properties [5]. Aloe species have antibacterial, antifungal, anticancer, antiviral and immunomodulatory properties [6]. *Aloe secundiflora* leaf components have been credited for antibacterial, antifungal and antiviral and anthelminthic medicinal properties [7]. *Aloe secundiflora* has been used in treating ailments including: chest problems, polio, malaria and stomach ache by herbalists in the Lake Victoria region in Kenya [8].

*Aloe secundiflora* leaf components have been credited for antibacterial, antifungal and antiviral and anthelminthic medicinal properties [9]. Aloe products have also been used in pharmaceuticals, cosmetic and food industries [10]. The main of the study was to provide insight about the antimicrobial activities of *Aloe secundiflora* leaf extracts and their use in treatment of bacterial or fungal infections.

Materials and Methods

Plant material collection

The fresh plant material of *Aloe secundiflora* was collected at Kenyatta University Arboretum. Voucher specimen was prepared and deposited in the university herbarium in Plant Sciences Department for future reference. The plants were brought to the laboratory and thoroughly washed in running water to remove debris and dust particles and then rinsed using distilled water and finally air dried.

Preparation of plant extract

The air dried plant materials were taise into powder and soaked in methanol for 72 hours, placed in a Gallenkamp shaker at 65 revolutions per minute. The contents were homogenized and filtered using whatman filter paper no. 1. The filtrate was poured into a round bottom flask and concentrated using a vacuum evaporator and stored in a labelled amber glass bottle at room temperature away from light and heat before being used for antimicrobial efficacy test.

Antimicrobial susceptibility testing

The microorganisms used were clinical isolates of *Escherichia coli*, *Salmonella typhi*, *Enterococcus faecalis*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Enterococcus faecalis*, *Staphylococcus aureus*, *Shigella flexneri*, *Enterococcus faecalis* and *Candida albicans* obtained from Kenyatta University Health Centre Laboratory, Nairobi. The test microorganisms were concentrated by comparing it with a 0.5 McFarland standard. Discs of 6 milliliters were prepared from whatman no.1 filter paper. The discs were sterilized by autoclaving. After sterilization the moisture discs were dried on hot air oven at 50°C [11]. The discs were impregnated with the extract from the highest concentration of 1000 mg/ml to the lowest concentration of 1 mg/ml by subsequently halving the dilutions [12]. Dimethyl sulphoxide (5%) was used in impregnating the extracts on the discs. Antimicrobial efficacy test was carried out using Kirby Bauer method [13]. Hektoen agar was used in the spread
plate technique where the clinical isolates were spread using sterilized cotton wool swabs. They were then exposed to discs impregnated with *Aloe secundiflora* leaf extract. The discs were placed with equal distance between them on agar plates inoculated with the bacterial pathogens and *Candida albicans*. Positive control discs containing ciprofloxacin was used for the bacteria's *Escherichia coli*, *Salmonella typhi*, *Enterococcus faecalis*, *Shigella flexineri*, *Staphylococcus aureus*, and fluconazole for *Candida albicans*. A negative control of discs impregnated with 5% DMSO was also used. The Petri dishes were incubated at 37°C for 24 hours.

The experiment was carried in duplicates and the average diameter of zones of inhibition formed determined. Minimal inhibitory concentration (MIC) was determined using microplate wells [14]. 100 µl of 250 mg/ml of methanol extract was added to 100 µl of sterile bacteriological peptone in the first well of the 96 well micro plate and mixed well with a micropipette. 100 µl of this dilution was transferred subsequently to wells two following each dilution of the original extract. An inoculum of 100 µl (0.5 McFarland standard) of overnight clinical cultures of; *Escherichia coli*, *Salmonella typhi*, *Staphylococcus aureus*, *Shigella flexineri*, *Enterococcus faecalis* and fungus *Candida albicans* were added in each of the wells. Triplicate of each micro plate were made and the procedure repeated for each of the test organisms. The plates were then incubated at 37°C for 24 hours. After incubation 40 µl of 0.2 mg/µl of INT was added in each of the wells and the plates examined after an additional 60 minutes of incubation. Growth was indicated by a red colour (conversion of INT to formazan). The lowest concentration at which the colour was apparently invisible as compared to the next dilution was taken as the minimum inhibitory concentration [15]. Minimum bacteroidal concentration (MBC) and minimum fungicidal concentration (MFC) was determined by taking 100 µl of suspension from micro plates wells that demonstrated no growth and inoculating on agar plates. The plates were incubated at 37°C for 24 hours. In the case where there was no bacterial growth and also not greater than the minimum inhibitory concentration was used to determine the maximum bacterial concentration and maximum fungicidal concentration [15].

Results

In this study the antimicrobial activity of *Aloe secundiflora* leaf was evaluated for antimicrobial activity against the selected human pathogens. Methanol was used in the extraction process and the antimicrobial activity determined using the Kirby Bauer method. Tables 1-3 shows the antimicrobial activity of the extract against bacterial pathogens and fungus *Candida albicans*. The results indicate that the extract from *Aloe secundiflora* was active against all the tested microorganisms. The positive control (ciprofloxacin) produced significantly sized zones of inhibition against the bacterial pathogens (ranging from 20-25 mm); fluconazole against *Candida albicans* (28 mm). Negative control dimethyl sulphoxide (DMSO) produced no zone of inhibition. The extract showed good antimicrobial activity against *Shigella flexineri* and *Enterococcus faecalis* producing highest zone of inhibition (18 mm) in both. The extract showed minimal antimicrobial activity (13 mm) against *Staphylococcus aureus*.

The MIC of the extracts ranged from 3-11 mg/ml while the MBC ranged from 7-13 mg/ml as indicated in Table 1. *Aloe secundiflora* extract was more active against *Shigella flexineri* and *Salmonella typhi* in low concentrations as compared to its activity against *Staphylococcus aureus*. The extract had a strong bactericidal capability against *Salmonella typhi* and *Shigella flexineri* in low concentrations (Table 3)

### Discussion

Medicinal plants have been used for a long time in traditional medicine to treat diseases. Over 122 compounds currently used in modern medicine have been derived from plants and 80% of their derivatives have had an ethno medicinal use [16]. The use of herbs to treat diseases is common among the non-industrialized societies where they are seen as more affordable than purchasing the modern pharmaceuticals [17]. The world health organization has estimated that 80 percent of population of Asia and African countries presently use herbal medicine for some of their primary health [17]. In this study, the antimicrobial potency of *Aloe secundiflora* was tested against Gram negative bacteria (*Escherichia coli*, *Salmonella typhi*, *Enterococcus faecalis* and *Shigella flexineri*) and fungal pathogen *Candida albicans*. The plant extract showed pronounced antimicrobial activity against *Salmonella typhi* and *Enterococcus faecalis* and less activity against *Staphylococcus aureus* (Table 1). When the extract was used in low concentrations, it showed pronounced inhibition activity against *Shigella flexineri* and *Salmonella typhi* when compared to *Staphylococcus aureus*. The antimicrobial activity of the *Aloe secundiflora* extract was less active against *Staphylococcus aureus* as compared to all the bacterial pathogens and fungal pathogen used. This showed that the extract from the plant had antimicrobial potential against the enteric bacterial pathogens and *Candida albicans*. Similar studies carried out have shown that extracts from *Aloe secundiflora* has antimicrobial potential against both fungal and bacterial pathogens [7,18]. Medicinal plants have known to produce secondary metabolites which have been thought to be responsible for the antimicrobial activity caused by their extracts. Some of the secondary metabolites such as alkaloids, polyphenols, glycosides and terpenes have shown antimicrobial activity against bacterial pathogens [19]. From the study, the plant extract from *Aloe secundiflora* contained saponins tannins, alkaloids and flavonoids. The phytochemicals present might be responsible for the antimicrobial activity. Tannin is a stringent vegetable product found in a wide range of plants parts ranging from the barks, roots, seeds, fruits, leaves, galls and roots [20]. Tannins are generally found in plants and they are thought to function as chemical defenses against pathogens and herbivory [21]. Tannins have shown to be responsible for inhibition of fungal and bacterial growth [22,23]. Flavonoids or bioflavonoids are secondary metabolites of plants that chemically have a general structure of 15 carbon skeleton consisting...

<table>
<thead>
<tr>
<th>Microorganisms</th>
<th>Extract</th>
<th>Ciprofloxacin</th>
<th>Fluconazole</th>
<th>DMSO</th>
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</thead>
<tbody>
<tr>
<td><em>Escherichia coli</em></td>
<td>17 ± 1.38 mm</td>
<td>20 ± 2.47 mm</td>
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<td>0.0 mm</td>
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<tr>
<td><em>Salmonella typhi</em></td>
<td>16 ± 0.68 mm</td>
<td>24 ± 0.35 mm</td>
<td>0.0 mm</td>
<td>0.0 mm</td>
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<tr>
<td><em>Staphylococcus aureus</em></td>
<td>13 ± 0.17 mm</td>
<td>25 ± 1.06 mm</td>
<td>0.0 mm</td>
<td>0.0 mm</td>
</tr>
<tr>
<td><em>Shigella flexineri</em></td>
<td>18 ± 0.38 mm</td>
<td>22 ± 1.06 mm</td>
<td>0.0 mm</td>
<td>0.0 mm</td>
</tr>
<tr>
<td><em>Enterococcus faecalis</em></td>
<td>18 ± 0.35 mm</td>
<td>22 ± 1.06 mm</td>
<td>0.0 mm</td>
<td>0.0 mm</td>
</tr>
<tr>
<td><em>Candida albicans</em></td>
<td>17 ± 0.38 mm</td>
<td>0.0 mm</td>
<td>28 ± 3.18 mm</td>
<td>0.0 mm</td>
</tr>
</tbody>
</table>

**Table 1:** Antimicrobial activity of *Aloe secundiflora* leaf extract against bacterial pathogens and fungus *Candida albicans*.
Table 2: Phytochemical analysis of leaf extract.

<table>
<thead>
<tr>
<th>Name of test</th>
<th>Saponins</th>
<th>Tannins</th>
<th>Alkaloids</th>
<th>Flavonoids</th>
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<tr>
<td>+</td>
<td>+</td>
<td>+</td>
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<td>+</td>
</tr>
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</table>

Key: (+) present

References


17. http://www.traffic.org/medicinal-plants


biosynthetic genes in herbaceous peony (Paeonia lactiflora Pall.). Electro J Biotech 15.


